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Serum Thymidine Kinase 1 – Potential Prostate Cancer Biomarker: A Clinical Study

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Serum Thymidine Kinase 1 – Potential Prostate Cancer Biomarker: A Clinical Study

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Abstract. *Background/Aim:* Serum thymidine kinase 1 (STK1) is a proliferation biomarker that has been used as a diagnostic marker of several malignant diseases. However, there are limited data for prostate cancer (PCa). *Patients and Methods:* In this study, we retrospectively analysed serum samples from 169 patients with biopsy confirmed PCa, who had been indicated for radical prostatectomy (RP) between 2013-2016. The results were compared with those in serum samples from 39 healthy men. We used commercially available enzymatic immunoassay to determine the levels of STK1. The patients were divided into groups according to the Gleason score (GS) and risk factors for adjuvant radiotherapy (aRT), which were defined as GS 8-10, pT3, and a positive surgical margin. *Results:* The median serum level of STK1 in PCa patients was 0.289 pmol/l. In the control group, the median value was 0.0116 pmol/l ($p < 0.001$). By comparing the patients with GS ≤ 6 vs. 7 vs. ≥ 8 ($p = 0.01$), we found statistically significant differences. In the correlation of STK1 values with risk factors, we found statistically significant differences both in comparison of 0 vs. 1 vs. 2 vs. 3 risk factors ($p = 0.021$), as well as ≤ 1 vs. ≥ 2 risk factors ($p = 0.009$). *Conclusion:* The levels of STK1 are significantly higher in patients with PCa than those in healthy controls. Furthermore, STK1 values correlate with GS and predefined risk factors for aRT. Therefore, STK1 can be considered as a potential tumour marker of PCa diagnosis and risk stratification.

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Key Words: Prostate cancer, thymidine kinase 1, STK1, serum biomarker.

Prostate cancer (PCa) is the second most common male malignancy, accounting for 14% of all diagnosed cancer cases with the incidence steadily growing (1). Currently, it is the third most common cause of cancer mortality among men, accounting for just over 10% of all cancer-related deaths (2). The prostate-specific antigen (PSA) is currently the most frequently used tumour marker in the early detection of PCa, despite its low specificity and low negative predictive value (3). False positives trigger unnecessary biopsies with a certain rate of complications. Furthermore, PCa represents a wide spectrum of diagnoses, ranging from clinically indolent to aggressive, high-grade cancers. PSA-based screening leads to overdiagnosis and overtreatment (4), escalating the overall cost of treatment. New markers could lead to better differentiation of significant cancer as well as improved monitoring of the disease. Therefore, research for novel cost-effective serum and urine diagnostic biomarkers with higher accuracy is needed.

Although almost one hundred potentially useful urine and serum markers for PCa have been reported, none of these have replaced PSA on its own or in a combination with other tumour markers (3, 5, 6). One of the main cancer characteristics is uncontrolled cell proliferation. Proliferative activity of cancer cells correlates with the aggressiveness of the disease. Predictive markers capable of measuring tumour-cell proliferation are clinically valuable because they may improve chances of early detection of tumour-related diseases, as well as its monitoring during therapy (7). Serum thymidine kinase 1 (STK1) is a proliferation biomarker that has already been used as a diagnostic marker for several malignant diseases (8).

STK1 is a cellular enzyme involved in the salvage pathway of DNA precursor synthesis. It catalyses the conversion of thymidine to deoxythymidine monophosphate, which is further phosphorylated to deoxythymidine di- and triphosphates prior incorporation into DNA (9).

Table I. *The characteristics of the patients and healthy subjects.*

Variable	Patients=169	Controls=39	p-Value
STK1, pmol/l; mean (SD; min - max)	0.289 (0.289; 0.062-1.78)	0.012 (0.0742; 0.0625-0.364)	<0.001
STK1 density, pmol/l/ml;			
mean (SD; min - max)	0.825 (0.902; 0.066-6.538)	X	X
iPSA, ng; mean (SD; min - max)	9.522 (7.929; 1.770-68.75)	X	X
iPSA density, ng/ml;			
mean (SD; min - max)	0.258 (0.208; 0.039-1.803)	X	X
Age, years; mean (SD; min - max)	64.928 (6.363; 40.9-79.1)	65.923 (5.441; 55-80)	0.475
Gleason score; n (%)			
6	68 (40)	X	X
7	79 (47)	X	X
8 and more	22 (13)	X	X
Stage; n (%)			
Localised (pT2)	110 (65)	X	X
Locally advanced	59 (35)	X	X
Surgical margin; n (%)			
Positive	47 (28)	X	X
Negative	122 (72)	X	X

STK1: Serum thymidine kinase 1; iPSA: prostate-specific antigen at the time of diagnosis; SD: standard deviation.

The activity of STK1 is cell-cycle dependent and shows a different pattern in normal proliferating cells compared with tumour cells. In normal cells, STK1 activity reaches its peak at late G1 phase/early S phase (10-20-fold increase) and is dramatically reduced to undetectable levels by the end of M phase (10, 11). However, STK1 activity may remain elevated in G2 and M phases of the cell cycle in malignant cells (12), most likely due to disordered regulation of transcription and degradation. High levels of STK1 have been observed in proliferating and malignant cells (13, 14).

In our pilot study, we measured increased concentrations of STK1 in patients with PCa (15). Our aim was to confirm these results in a larger study. We believe that STK1 may prove to be a cost-efficient and minimally invasive diagnostic and monitoring tool for PCa.

Patients and Methods

Patient selection. We performed a retrospective analysis of prospectively collected serum samples from 169 patients with PCa scheduled for radical prostatectomy (RP) between 2013-2016. The results were compared with those in the serum from 39 healthy male volunteers, with an average age of 61 years, without relevant urological or oncological medical history, with negative urine bacterial culture and urine cytology, PSA level under 2 µg/l, and negative digital rectal examination. The Institutional Ethics Committee approved the study. All participants gave their written informed consent.

Sample and data collection. Blood was collected from all patients before RP after overnight fasting via puncture of the cubital vein. The blood was then centrifuged for 10 min at 3,000 rpm (1,450 g) and the serum was aliquoted, immediately frozen, and kept at -70°C

until STK1 was analysed. Radical prostatectomy specimens were examined by a specialized pathologist. Tumours were classified according to the tumour, node and metastasis (TNM) classification (16) and graded according to the Gleason score (GS) (17). Healthy volunteers underwent the same blood sampling procedure.

Concentration measurements. For both groups, we measured the STK1 marker level in the serum with the use of enzyme-linked immunosorbent assay (ELISA), utilizing a commercially available immunoassay technique ELISA kit (LSBio, Inc, Seattle, WA, USA).

The analytical parameters of the kit were as follows: detection limit 0.063 pmol/l and working range 0.063-4.0 pmol/l [intra-assay coefficient of variation (CV)=5.3%/inter-assay CV=8.6%]. Serum PSA levels were measured using the electrochemiluminescence sandwich immunoassay on the Cobas e6000 analyser (Hitachi, High Technology Corp., Tokyo, Japan).

For statistical analysis, the patients were divided into groups in accordance with GS and risk factors for adjuvant radiotherapy (aRT), which were defined as GS 8-10, pT3 or a positive surgical margin.

Statistical analysis. Statistical data analysis was performed using SAS software (SAS Institute Inc., Cary, NC, USA). Basic statistical data such as mean, standard deviation, variance, median, interquartile range, minimum and maximum were calculated for the measured parameters. For categorical variables, their absolute and relative frequencies were examined. Nonparametric tests (Wilcoxon's two-sample test and its generalised variant, the so-called Kruskal-Wallis test) were used to compare the distributions of the examined parameters between the tested groups. We tested the age agreement between the examined groups using Two One Sample Test. The relationships between the parameters were investigated using the Spearman correlation coefficient and were expressed graphically using linear regression. Statistical significance was determined at alpha=5%.

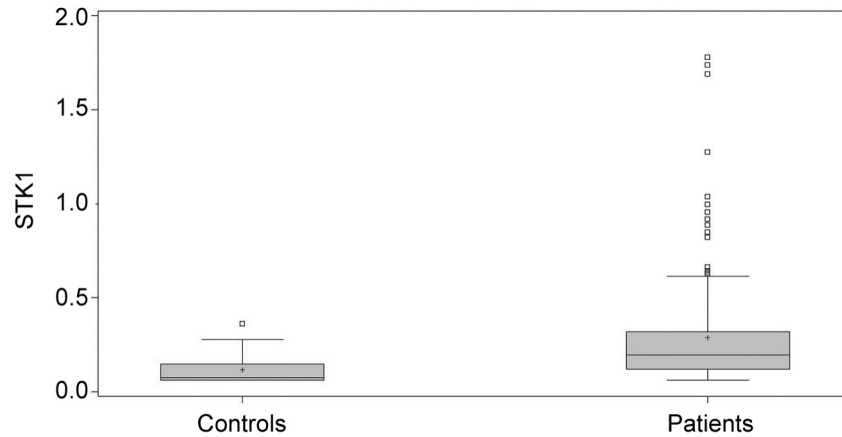


Figure 1. Levels of serum thymidine kinase 1 (STK1) (pmol/l) in patients with confirmed prostate cancer and healthy controls.

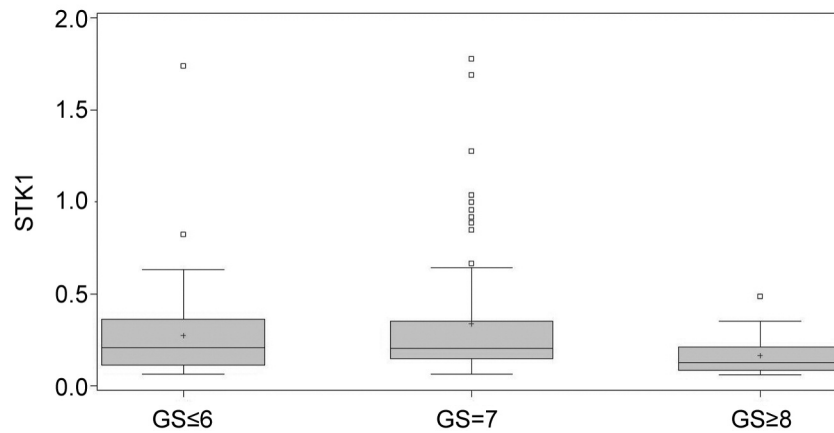


Figure 2. Correlation between Gleason score and levels of serum thymidine kinase 1 (STK1) (pmol/l).

Results

The study included 169 patients with PCa aged between 40 and 79 years and 39 healthy subjects aged between 53 and 78 years. The characteristics of the patients and healthy subjects are summarized in Table I.

The serum levels of STK1 in PCa patients were significantly increased as compared to those in the control group of healthy subjects. The median STK1 level detected in PCa patients was 0.289 pmol/l [standard deviation (SD)=0.289; min-max 0.062-1.78], whereas in the control group was 0.012 pmol/l (SD=0.0742; min-max 0.063-0.364) ($p<0.001$) (Figure 1, Table I).

When comparing patients with GS≤6 vs. 7 vs. ≥8 ($p=0.01$) (Figure 2, Table II), we found a statistically significant difference, but not when comparing patients with GS≤6 vs.

≥7 ($p=1.000$). Staging did not show any significant difference neither in the comparison of pT2 vs. pT3a vs. pT3b ($p=0.989$), nor in the comparison of pT2 vs. pT3a+pT3b ($p=1.0$) (Table II).

We found a statistically significant correlation of STK1 serum level with risk factors for aRT (GS 8-10, pT3, positive surgical margin), $p=0.021$ (Figure 3). The lowest levels of STK1 were found in patients with all three risk factors. The difference between patients with no risk factors and to those with at least one risk factor was not significant ($p=0.579$). Statistically significantly lower levels of STK1 were found when comparing patients with 0 or 1 and patients with 2 or more risk factors ($p=0.009$) (Table III, Figure 4).

Correlations between STK1 and PSA density, PSA at the time of diagnosis, prostate health index (PHI), and prostate size measured by TRUS were not statistically significantly different.

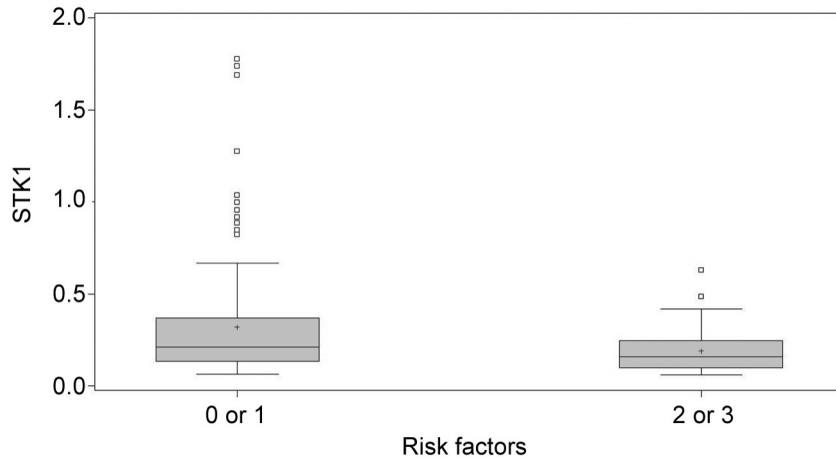


Figure 3. Correlation between number of risk factors for adjuvant radiotherapy (Gleason score 8-10, pT3, positive surgical margin) and levels of serum thymidine kinase 1 (STK1) (pmol/l).

Table II. Correlation between staging and levels of STK1.

Parameter	STK1, pmol/l; median (SD; min-max)	p-Value
GS≤6	0.21 (0.25; 0.063-1.74)	0.098
GS7	0.204 (0.34; 0.062-1.78)	
GS≥8	0.127 (0.111; 0.062-0.49)	1.0
GS≤6	0.21 (0.25; 0.063-1.74)	
GS≥7	0.193 (0.31; 0.062-1.78)	0.989
pT2	0.594 (0.862; 0.066-5.235)	
pT3a	0.531 (1.121; 0.134-6.538)	1
pT3b	0.715 (0.467; 0.17-1.633)	
pT2	0.594 (0.862; 0.066-5.235)	
pT3a+pT3b	0.535 (0.985; 0.134-6.538)	

STK1: Serum thymidine kinase 1; SD: standard deviation; GS: Gleason score.

Table III. Correlation between risk factors for adjuvant radiotherapy (GS 8-10, pT3, positive surgical margin) and levels of STK1.

Risk factors	STK1, pmol/l; median (SD; min-max)	p-Value
0	0.196 (0.342; 0.063-1.78)	0.0211
1	0.265 (0.271; 0.063-1.278)	
2	0.16 (0.124; 0.067-0.634)	
3	0.084 (0.085; 0.062-0.243)	0.579
0	0.196 (0.342; 0.063-1.78)	
1 or 2 or 3	0.191 (0.224; 0.062-1.278)	0.792
0	0.196 (0.342; 0.063-1.78)	
2 or 3	0.195 (0.226; 0.063-1.278)	0.009
0 or 1	0.21 (0.319; 0.063-1.78)	
2 or 3	0.158 (0.122; 0.062-0.634)	

STK1: Serum thymidine kinase 1; GS: Gleason score.

Discussion

We compared serum levels of STK1 in patients with PCa and healthy controls using an enzyme immunoassay method utilising a commercially available kit. We showed that serum levels of STK1 in patients with PCa were significantly higher (median 0.289 pmol/l) than those in the control group (median 0.0116 pmol/l).

While PSA will likely remain the most widely used prostate tumour marker in the near future, the need for other diagnostic methods, either on their own or in combination with other tumour markers, is becoming more and more urgent, in particular because of the sensitivity and specificity of the PSA test. The potential new marker must fulfil several

requirements such as cost efficiency, minimal invasiveness and repeatability (18).

The results of our study confirm our hypothesis that higher STK1 level is associated with the diagnosis of PCa. The presented results are in accord with the study of Li *et al.* (19), who demonstrated that STK1 concentration and total PSA were significantly higher in patients with PCa, as compared to patients with benign prostatic hyperplasia (BPH) and healthy individuals [n=123, median 2.5 pmol/l (SD 2.0; min-max 0.2-14.7)]. Furthermore, STK1 concentration was associated with GS, whereas total PSA was not. However, no association was identified between STK1 concentration and total serum PSA. Li *et al.* also indicated the difficulty in differentiating BPH from PCa, *i.e.*, supporting our aim for the identification of markers

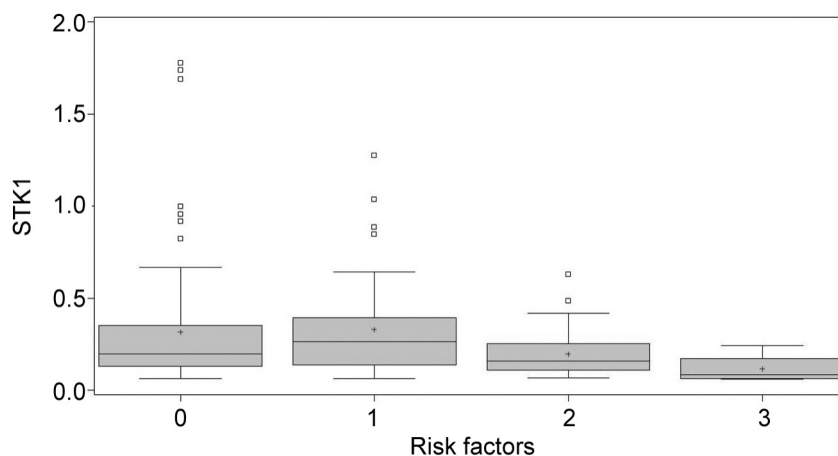


Figure 4. Correlation between number of risk factors for adjuvant radiotherapy (Gleason score 8-10, pT3, positive surgical margin) and levels of serum thymidine kinase 1 (STK1) (pmol/l).

to be used in a combination with other molecules. Our results also show the association of STK1 concentrations with the grading of the disease according to GS.

Lundgren *et al.* (20) measured STK1 in 36 patients, who died of PCa [median 0.30 ng/ml (min-max 0.21-0.41)], and in 294 randomly selected healthy men, and showed that high levels of STK1 can predict PCa-related death in 30 years. Another study by Jagarlamudi *et al.* (8) compared STKa (serum thymidine kinase activity) and concentration of STK1 in patients with PCa (n=47) to those of healthy blood donors. The results demonstrated that STKa and STK1 concentration differed significantly between patients with PCa and healthy individuals.

In this study, we measured the levels of STK1 in a higher number of patients with PCa than in previous studies. In addition, unlike others, we added risk factors for aRT after RP (GS 8-10, pT3, positive surgical margin), which may prove as a useful predictive combination of parameters with clinical benefits. Our results confirm that STK1 levels are significantly higher in patients with PCa than in healthy controls. Furthermore, STK1 values correlate with the tumour GS and a number of predefined risk factors for aRT. Therefore, STK1 is a promising tumour marker for PCa. It is important to note that the current study was not a clinical trial following specific criteria, rather, it was based on data collected during routine clinical practice. This may limit the reliability of the conclusions drawn.

In the future, we plan to examine a panel of serum and urine biomarkers which could function as a reliable guidance for PCa diagnosis with higher sensitivity and specificity than PSA. In turn, the number of unnecessary prostate biopsies, which are both invasive and pose significant risks for the patients, will be reduced.

Conflicts of Interest

The Authors have no conflicts of interest to report in relation to this study.

Authors' Contributions

Rezac Jakub drafted the manuscript and prepared the figures. Measurements were performed by Hanouskova Lenka, and Kotaska Karel. Kantorova Alzbeta, Linhartova Anna and Fiala Vojtech edited the manuscript. Supervision was performed by Capoun Otakar, Soukup Viktor, and Vesely Stepan.

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References

- 1 Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A and Bray F: Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 71(3): 209-249, 2021. PMID: 33538338. DOI: 10.3322/caac.21660
- 2 Health at a Glance 2019: OECD Indicators, 2019. Available at: <https://www.oecd-ilibrary.org/sites/ac388762-en/index.html?itemId=/content/component/ac388762-en> [Last accessed on August 30, 2022]
- 3 Filella X and Foj L: Emerging biomarkers in the detection and prognosis of prostate cancer. *Clin Chem Lab Med* 53(7): 963-973, 2015. PMID: 25581761. DOI: 10.1515/cclm-2014-0988

- 4 Dani H and Loeb S: The role of prostate cancer biomarkers in undiagnosed men. *Curr Opin Urol* 27(3): 210-216, 2017. PMID: 28212119. DOI: 10.1097/MOU.0000000000000384
- 5 Bratt O and Lilja H: Serum markers in prostate cancer detection. *Curr Opin Urol* 25(1): 59-64, 2015. PMID: 25393274. DOI: 10.1097/MOU.0000000000000128
- 6 Shariat SF, Semjonow A, Lilja H, Savage C, Vickers AJ and Bjartell A: Tumor markers in prostate cancer I: blood-based markers. *Acta Oncol* 50(Suppl 1): 61-75, 2011. PMID: 21604943. DOI: 10.3109/0284186X.2010.542174
- 7 He Q, Fornander T, Johansson H, Johansson U, Hu GZ, Rutqvist LE and Skog S: Thymidine kinase 1 in serum predicts increased risk of distant or loco-regional recurrence following surgery in patients with early breast cancer. *Anticancer Res* 26(6C): 4753-4759, 2006. PMID: 17214336.
- 8 Jagarlamudi KK, Hansson LO and Eriksson S: Breast and prostate cancer patients differ significantly in their serum Thymidine kinase 1 (TK1) specific activities compared with those hematological malignancies and blood donors: implications of using serum TK1 as a biomarker. *BMC Cancer* 15: 66, 2015. PMID: 25881026. DOI: 10.1186/s12885-015-1073-8
- 9 Eriksson S, Munch-Petersen B, Johansson K and Eklund H: Structure and function of cellular deoxyribonucleoside kinases. *Cell Mol Life Sci* 59(8): 1327-1346, 2002. PMID: 12363036. DOI: 10.1007/s00018-002-8511-x
- 10 Jagarlamudi KK and Shaw M: Thymidine kinase 1 as a tumor biomarker: technical advances offer new potential to an old biomarker. *Biomark Med* 12(9): 1035-1048, 2018. PMID: 30039979. DOI: 10.2217/bmm-2018-0157
- 11 Sherley JL and Kelly TJ: Regulation of human thymidine kinase during the cell cycle. *J Biol Chem* 263(17): 8350-8358, 1988. PMID: 3372530.
- 12 Chang ZF and Huang DY: The regulation of thymidine kinase in HL-60 human promyeloleukemia cells. *J Biol Chem* 268(2): 1266-1271, 1993. PMID: 8419329.
- 13 He Q, Skog S and Tribukait B: Cell cycle related studies on thymidine kinase and its isoenzymes in Ehrlich ascites tumours. *Cell Prolif* 24(1): 3-14, 1991. PMID: 2009315. DOI: 10.1111/j.1365-2184.1991.tb01506.x
- 14 Gasparri F, Wang N, Skog S, Galvani A and Eriksson S: Thymidine kinase 1 expression defines an activated G1 state of the cell cycle as revealed with site-specific antibodies and ArrayScan assays. *Eur J Cell Biol* 88(12): 779-785, 2009. PMID: 19726104. DOI: 10.1016/j.ejcb.2009.06.005
- 15 Hanousková L, Řezáč J, Veselý Š, Průša R and Kotaška K: Thymidine kinase-1 as additional diagnostic marker of prostate cancer. *Clin Lab* 66(6), 2020. PMID: 32538039. DOI: 10.7754/Clin.Lab.2019.191026
- 16 The European Association of Urology (EAU) prostate cancer guidelines, 2022. Available at: <https://uroweb.org/guidelines/prostate-cancer> [Last accessed on August 30, 2022]
- 17 Gleason DF: Classification of prostatic carcinomas. *Cancer Chemother Rep* 50(3): 125-128, 1966. PMID: 5948714.
- 18 Biomarkers in cancer: an introductory guide for advocates. Available at: <https://cancer.wisc.edu/research/wp-content/uploads/2019/05/Biomarkers-in-Cancer.pdf> [Last accessed on December 15, 2022]
- 19 Li S, Zhou J, Wang Y, Zhang K, Yang J, Zhang X, Wang C, Ma H, Zhou J, He E and Skog S: Serum thymidine kinase 1 is associated with Gleason score of patients with prostate carcinoma. *Oncol Lett* 16(5): 6171-6180, 2018. PMID: 30333882. DOI: 10.3892/ol.2018.9345
- 20 Lundgren PO, Tribukait B, Kjellman A, Norming U, Jagarlamudi K and Gustafsson O: Serum thymidine kinase 1 concentration as a predictive biomarker in prostate cancer. *Prostate* 82(8): 911-916, 2022. PMID: 35294068. DOI: 10.1002/pros.24335

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ORIGINAL ARTICLE

Thymidine Kinase-1 as Additional Diagnostic Marker of Prostate Cancer

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SUMMARY

Background: Thymidine kinase-1 (TK-1) is associated with proliferation and malignancy and has been extensively studied as a diagnostic biomarker for a variety of tumors, but there are limited data for prostate cancer.

Methods: TK-1 concentrations in serum were measured in 59 patients with prostate cancer (mean age 68 years) and in the control group of 28 healthy men (mean age 63 years) using commercially available enzymatic immuno-assay (LSBio, Inc., Seattle, WA, USA). The patients were divided with respect to the severity of the disease into two groups according to the European Association of Urology (EAU) guidelines (Stage 1, 2 - less severe tumors, stage 3 - severe tumors).

Results: Serum thymidine kinase-1 concentrations were significantly elevated in the group of the patients with prostate cancer compared to the healthy individuals (0.204 pmol/L vs. 0.072 pmol/L, with $p < 0.0001$). Diagnostic efficiency of serum TK-1 concentrations was 0.792 with the specificity of 53.6% and sensitivity of 94.9%. Patients with less severe tumors (Stage 1, 2) and severe tumors (Stage 3) had significantly increased levels of TK-1 as well ($p < 0.0001$). Combination of TK-1 and PSA investigation in patients with PCa improve the diagnostic validity of TK-1 (AUC = 0.87).

Conclusions: Concentrations of thymidine kinase 1 are increased in all patients with prostate cancer and even more in patients with severe prostate cancer. Thymidine kinase 1 appears to be a promising additional diagnostic marker promising in patients with prostate cancer.

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KEY WORDS

prostate cancer, thymidine kinase-1

INTRODUCTION

Prostate cancer (PCa) is one the most frequent malignancy of men and the third most common cause of death in Czech Republic men [1]. Currently, prostate-specific antigen (PSA) is the most widely used conventional serum marker including serum free PSA and fPSA/tPSA ratio. PSA velocity, PSA density, [-2] pro-PSA, and prostate health index (PHI) improve specificity, but values of these biomarkers might be affected by many processes [1,2]. Despite all of these markers, 100% conclusively precise diagnostic test for PCa has not yet been introduced. Thymidine kinase is an en-

zyme, a phosphotransferase (a kinase): 2'-deoxythymidine kinase, ATP-thymidine 5'-phosphotransferase. It is present in two forms in mammalian cells, TK-1 and TK-2. Two human TK genes encode two isoenzymes. The first form, TK-1, is located on chromosome 17q-25.3. TK-1 is present in the cytoplasm of cells and TK-1 is dependent on cell cycle. The second form is mitochondrial enzyme TK-2. This form is located on chromosome 16q21 and is cell cycle-independent [3,4]. TK-1 has been extensively studied as a diagnostic biomarker for a variety of cancer types, because TK-1 is the biomarker of proliferation and is involved in the pathway of DNA precursor synthesis [5-7]. TK-1 has been expressed in proliferating and malignant cells [6,8]. Elevated TK-1 activities or concentrations have been found in many tumors including gastric, ovary, cervical, esophageal, lung, prostate, and breast cancers [9]. The aim of our study was to investigate serum levels of TK-1 as a potential diagnostic biomarker in patients with prostate cancer.

MATERIALS AND METHODS

The serum samples of patients with prostate cancer were obtained in the morning before prostatectomy. The cancer diagnosis was performed by histological examination of tumor specimens obtained by prostate resection. The samples of patients and healthy individuals were aliquoted and frozen immediately and kept at -70°C until TK-1 was analyzed. Serum concentrations of TK-1 were measured in 59 patients (mean age 68, range 45 - 82 years) with prostate cancer, and the control group consisting of 28 healthy men (mean age 63 years, range 54 - 78 years) with non-malignant etiology of the disease including benign prostate hyperplasia. All subjects were informed about the project and signed an informed consent. The characteristics of the patients and healthy subjects are summarized in Table 1. Serum TK-1 levels were assayed using a commercially available immunoassay ELISA kit (LSBio, Inc., Seattle, WA, USA). The analytical parameters of the kit were: detection limit 0.063 pmol/L and working range 0.063 -4.0 pmol/L (intra-assay CV = 5.3%/inter-assay CV = 8.6%). Serum PSA levels were assayed using the electrochemiluminescence sandwich immunoassay on the Cobas e6000 analyzer (Hitachi, High Technology Corp., Tokyo, Japan). The patients were previously clinically investigated and classified according to the TNM classification [3]. For further investigation, the patients were divided into two groups according to the severity of the disease. The first group consisted of patients with less severe tumors (Stage 1 and 2), and the second group consisted of patients with severe tumors (Stage 3).

Statistical analysis

The differences between the subgroups were tested for the statistical significance by the nonparametric Mann-

Whitney test. A value of $p < 0.005$ was considered statistically significant. Receiving operating curve (ROC) analysis was used to examine the diagnostic efficiency. Analysis of variance was used to evaluate the correlation of TK-1 levels with age and PSA levels. The statistical software MedCalc version: 18.02.01 (Ostende, Belgium) was used for the statistical analysis.

RESULTS

The serum levels of thymidine kinase-1 in patients with prostate cancer were significantly increased compared with the control group of healthy men (median = 0.204 pmol/L vs. median = 0.072 pmol/L, $p < 0.0001$, Mann-Whitney test, Figure 1). Diagnostic efficiency of serum TK-1 expressed as AUC calculated from the ROC analysis was 0.792 (specificity = 53.6% and sensitivity = 94.9%) (Figure 2). The serum levels of TK-1 in patients with pT1, 2 and pT3 stages were significantly increased compared with the control group ($p < 0.0001$ and $p = 0.0026$, respectively, one-way analysis of variance) (Figure 3). The AUC for the combination of TK-1 and PSA is higher than those for the individual TK-1 (AUC = 0.87 vs. AUC = 0.79, $p = 0.19$) (Figure 4).

DISCUSSION

The results of the pilot study confirmed the relevant role of TK-1 in the prostate cancer diagnosis. We proved that the serum levels of TK-1 were significantly increased in the patients with prostate cancer compared with the healthy individuals. These results correlate with results of other studies [6,8]. These findings correlated with the results previously published in study of Jagarlamundi et al. [6] with AUC = 0.88, sensitivity = 0.64, and specificity = 0.96. The presented study showed very good diagnostic efficiency of serum TK-1 values (AUC = 0.79, with specificity 54% and sensitivity 95%). The differences in AUC found in our study and in study of Jagarlamundi et al. are not significant ($p = 0.14$). These results show that investigation of serum TK-1 levels indicate that the determination of TK-1 concentration in serum might be a useful test even for the screening of individuals for prostate cancer risk. Recent publications mentioned the investigation of catalytic activity of TK-1 with respect to the tumor proliferation and progression; nevertheless, the diagnostic power of the TK-1 concentration seems to be higher than for the TK-1 catalytic activity. TK-1 concentrations show greater sensitivity for the solid tumors and show more consistent TK-1 values in different disease types [7]. We found significant differences between serum TK-1 concentrations in control group of healthy individuals and patients according to the pT1 and 2, and pT3 clinical stages ($p < 0.0001$ and $p = 0.0026$, respectively). These results correlate with the fact, that TK-1 expres-

Table 1. Characteristics of patients and healthy subjects.

	Control group (n = 28)	Patients (n = 59)	p-value
Mean age (range)	64 (55 - 78)	68 (45 - 82)	-
S-TK-1 (pmol/L)	0.072 (0.063 - 0.364)	0.204 (0.063 - 1.40)	< 0.0001
PSA (µg/L)	1.25 (0.29 - 2.34)	7.32 (3.02 - 19.27)	< 0.0001

Age expressed as mean (min - max); concentrations S-Thymidine kinase-1 and PSA are expressed as median (min - max), p-value is calculated with the Mann-Whitney test.

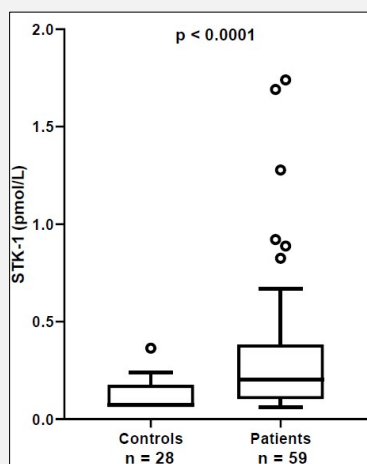


Figure 1. Serum TK-1 levels in healthy individuals and patients with prostate cancer (pmol/L).

Results are expressed as box-and-whisker plots with medians (IQR, 25th - 75th percentiles).

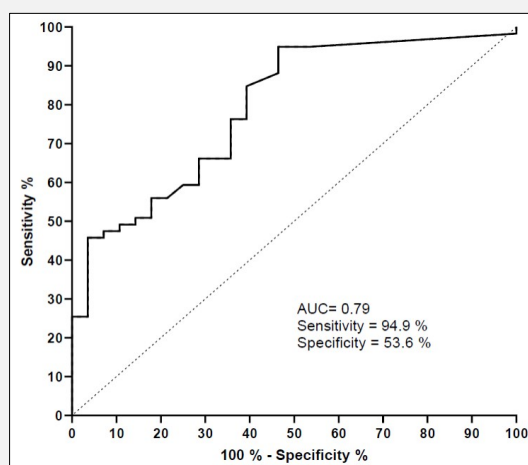


Figure 2. Diagnostic efficiency of TK-1.

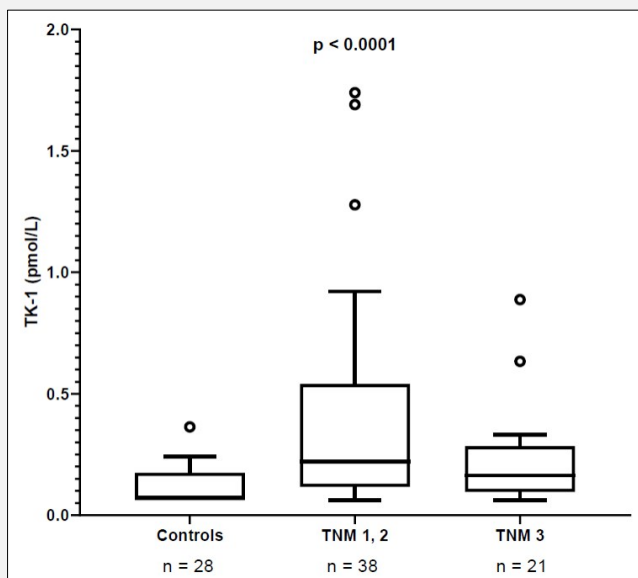


Figure 3. Serum TK-1 levels in patients with less severe and severe prostate cancer tumors.

Results are expressed as box-and-whisker plots with medians (IQR, 25th - 75th percentiles).

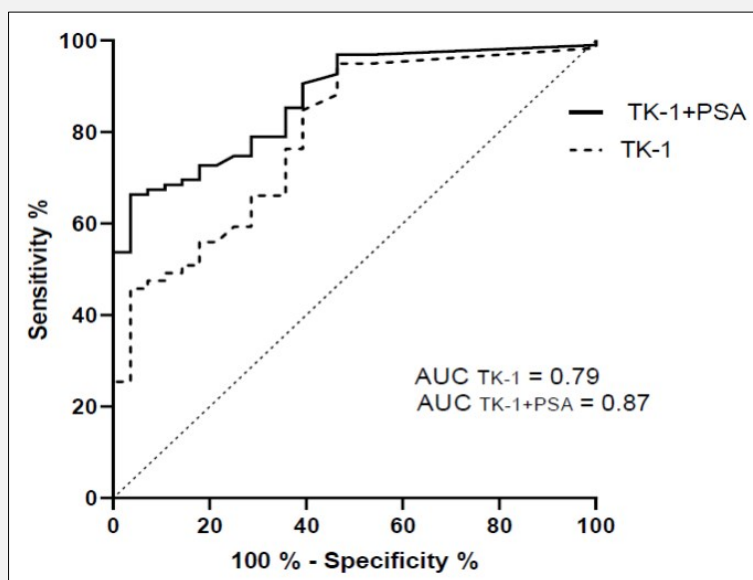


Figure 4. The diagnostic performance of TK-1 and PSA in patients with CaP.

sion is mostly associated with tumor proliferation. Recent publications also show the association of TK-1 concentrations with the grading of the disease according to the Gleason score (GS). We found significant differences between control group of healthy individuals and patients with GS = 5 + 6 and GS = 7 ($p = 0.002$ and $p < 0.0001$, respectively) as well. The results of our study were consistent with those published by Jagarlamudi et al. and Li et al. [6,8], which demonstrated that TK-1 levels correlate with the Gleason score. We also found that serum TK-1 levels are not significantly associated with PSA levels and age ($p = 0.41$, analysis of variance). We also confirmed that the AUC for the combination of TK-1 and PSA is higher than for the individual TK-1 (AUC = 0.87 vs. AUC = 0.79, $p = 0.19$). This finding lead to the opinion, that the combination of S-TK-1 and PSA investigation in patients with PCa improved the diagnostic validity of TK-1.

We also investigated other possible prostate cancer biomarkers (endoglin, TIMP-1, SPINK-1, chromogranin A and annexin A3), but none of them showed significant diagnostic power compared to TK-1 (AUC varied from 0.51 to 0.62). Our previous study confirmed that investigation of mindin levels in serum seems to be relevant for the diagnosis of prostate cancer. The concentrations of mindin in patients with prostate cancer are significantly decreased compared with the control group with AUC = 0.70 and are also correlated with the Gleason score and the staging of the cancer [10].

The presented study is just a pilot study, and our results need to be confirmed with a larger number of samples. It is important to emphasize, that TK-1 concentrations are assay dependent.

CONCLUSION

Concentrations of thymidine kinase-1 in serum are increased in patients with prostate cancer. They are not significantly related to the age and PSA levels. TK-1 appears to be a promising additional diagnostic marker useful in the diagnosis of prostate cancer.

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Declaration of Interest:

The authors state that there are no conflicts of interest regarding the publication of this article.

References:

1. Geryk E, Dítě P, Kozel J, et al. Other primary neoplasm in patients with prostate cancer in comparison of its incidence, mortality and prevalence. *Onkologie* 2010;4(2):89-94. <https://www.onkologics.cz/pdfs/xon/2010/02/08.pdf>
2. Ayyildiz SN, Ayyildiz A. PSA, PSA derivatives, proPSA and prostate health index in the diagnosis of prostate cancer. *Turk J Urol* 2014;40(2):82-8 (PMID: 26328156).
3. Heidenreich A, Aus G, Bolla M, et al. EAU guidelines on prostate cancer. *Eur Urol* 2008;53(1):68-80 (PMID: 17920184).
4. Aufderklamm S, Todenhofer T, Gakis G, et al. Thymidine kinase and cancer monitoring. *Cancer Lett* 2012;316(1):6-10 (PMID: 22068047).
5. Karlstrom AR, Neumuller M, Gronowitz JS, Kallander CF: Molecular forms in human serum of enzymes synthesizing DNA precursors and DNA. *Mol Cell Biochem* 1990;92(1):23-35 (PMID: 2155379).
6. Jagarlamudi KK, Hansson LO, Eriksson S. Breast and prostate cancer patients differ significantly in their serum Thymidine kinase 1 (TK1) specific activities compared with those hematological malignancies and blood donors: implications of using serum TK1 as a biomarker. *BMC Cancer* 2015;15:66 (PMID: 25881026).
7. Jagarlamudi KK, Shaw M. Thymidine kinase 1 as a tumor biomarker: technical advances offer new potential to an old biomarker. *Biomark Med* 2018, 12(9):1035-48 (PMID: 30039979).
8. Li S, Zhou J, Wang Y, Zhang K, et al. Serum thymidine kinase 1 is associated with Gleason score of patients with prostate carcinoma. *Oncol Lett* 2018;16(5):6171-80 (PMID: 30333882).
9. Xiang Y, Zeng H, Liu X, et al. Thymidine kinase 1 as a diagnostic tumor marker is of moderate value in cancer patients: A meta-analysis. *Biomed Rep* 2013;1(4):629-37 (PMID: 24648999).
10. Hanouskova L, Řezáč J, Veselý Š, et al. Diagnostic benefits of mindin as a prostate cancer biomarker. *Journal of Medical Biochemistry* 2019 (Ahead of Print). <https://content.sciendo.com/view/journals/jomb/ahead-of-print/article-10.2478-jomb-2019-0008.xml>

DIAGNOSTIC BENEFITS OF MINDIN AS A PROSTATE CANCER BIOMARKER DIJAGNOSTIČKE PREDNOSTI MINDINA KAO BIOMARKERA RAKA PROSTATE

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Summary

Background: It has been shown that decreased expression and activity of extracellular matrix protein mindin correlate with various types of cancers including breast, colon and lung cancers. The aim of the presented study was to investigate the serum mindin levels in prostate cancer.

Methods: Mindin concentrations in serum were measured in 56 patients with prostate cancer (mean age 68 years) and in control group of 29 healthy men (mean age 64 years) using commercially available enzymatic immunoassay (Cusabio, WuHan, China). The patients were divided with respect to the severity of the disease into two groups according to the EAU guidelines (stage 1, 2 – less severe tumours, stage 3, 4 – severe tumours).

Results: Serum mindin concentrations were significantly elevated in the group of healthy individuals unlike in the patients with prostate cancer (2.12 ng/mL vs 0.78 ng/mL, with $P=0.0007$, $AUC=0.705$). Patients with less severe tumours (stage 1, 2) and severe tumours (stage 3, 4) had significantly decreased levels of S-mindin as well ($P=0.0037$), although the difference in serum mindin concentrations between the patients with less severe and severe tumours was not significant.

Conclusions: Concentrations of mindin were decreased in patients with prostate cancer and reduced in patients with less severe prostate cancer as well. Mindin appears to be a promising diagnostic marker useful in the diagnosis of prostate cancer.

Keywords: mindin, prostate cancer, biomarker

Kratka sadržaj

Uvod: Pokazalo se da je smanjena ekspresija i aktivnost ekstracelularnog proteinskog matriksa mindina korelira sa različitim tipovima raka, uključujući rak dojke, debelog creva i pluća. Cilj prikazane studije je bio da se ispita serumski nivo mindina kod karcinoma prostate.

Metode: Koncentracije mindina u serumu su merene kod 56 bolesnika sa karcinomom prostate (srednja starost 68 godina) i u kontrolnoj grupi od 29 zdravih muškaraca (srednja starost 64 godine) korištenjem komercijalno dostupnog enzimskog testa (Cusabio, WuHan, Kina). Pacijenti su prema težini bolesti podeljeni u dve grupe prema EAU smernicama (faza 1, 2 – manje teški tumori, faza 3, 4 – teški tumori).

Rezultati: Koncentracije serumskog mindina bile su značajno povišene u grupi zdravih pojedinaca, za razliku od pacijenata sa rakom prostate (2,12 ng/mL u odnosu na 0,78 ng/mL, sa $P=0,0007$, $AUC=0,705$). Pacijenti sa manje ozbiljnim tumorima (faza 1, 2) i teškim tumorima (faza 3, 4) imali su i značajno snižene nivoe S-mindina ($P=0,0037$), iako razlika u koncentracijama mindina u serumu između pacijenata sa manje teškim i ozbiljnim tumorima nije bila značajna.

Zaključak: Koncentracije mindina su smanjene kod pacijenata sa rakom prostate, a smanjene su i kod pacijenata sa manje teškim oblikom raka prostate. Izgleda da je mindin obećavajući dijagnostički marker i da je koristan u dijagnostici raka prostate.

Ključne reči: mindin, rak prostate, biomarker

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List of abbreviations: AUC, Area under the curve; CV, Coefficient of variation; EAU, European Association of Urology; ELISA, Enzyme-linked immunosorbent assay; IQR, interquartile range; Pca, prostate carcinoma; PHI, prostate health index; PSA, Prostate-specific antigen; PSADT, Prostate-specific antigen doubling time; ROC, Receiver operating characteristics.

Introduction

Prostate cancer (PCa) is one the most frequently diagnosed tumours of men and the third most common cause of cancer death of Czech men (1). Currently, the only biomarker in wide clinical use for the diagnosis and prognosis of prostate cancer is the prostate-specific antigen (PSA). Other markers useful for the diagnostics of prostate cancer are free PSA and Free/Total PSA ratio, complexed PSA, pro PSA - 2proPSA and Prostate Health Index (PHI). Total PSA, % fPSA, f/tPSA, and other PSA derivatives, PSAD, PSA velocity, PSADT, age-specific PSA do not decrease the number of unnecessary biopsies performed for diagnostic purposes. Despite all of these markers, a 100% conclusively precise diagnostic test for PCa has not been introduced yet (2).

PSA can be detected in the serum of a blood sample and is considered to be currently the most useful tumour marker (3). PSA can be used for prostate cancer screening and monitoring of the response to the treatment. PSA seems to be useful for the detection of prostate cancer of the men whose total PSA concentration in the 4–10 mg/L range (4). Investigation of the serum PSA levels does not have a direct correlation with increasing grade and stage of prostate cancer (5). 20 % of patients have PSA levels less than 4 mg/L, 25 % of the patients have PSA levels in the interval of 4–10 mg/L. Moreover, some aggressive forms of prostate cancer can be PSA negative (6). Due to the limitations of PSA as a biomarker, there is still a need for new biomarkers that can be used as prognostic indicators of prostate cancer for effective differentiation between indolent and aggressive disease (7).

Mindin, also called spondin 2, is an extracellular matrix protein which is encoded by the SPO2 gene located in the chromosome 4p16.3. Spondin 2 belongs to the F-spondin family of secreted extracellular matrix proteins. The members of F-spondin family have three domains: FS1 (for F-spondin), FS, and thrombospondin type 1 repeats. Mindin exerts a broad spectrum of effects on the innate immune system and its role in cancer is currently investigated (8). Recently, mindin is mentioned as a candidate biomarker for prostate cancer diagnosis (9).

The study aimed to investigate serum levels of mindin as a potential diagnostic biomarker in patients with prostate cancer.

Materials and Methods

Serum samples of patients with prostate cancer were obtained in the morning before prostatectomy. The cancer diagnosis was performed by histological examination of tumour specimens obtained by prostate resection. The samples of patients and healthy individuals were frozen immediately, aliquoted and

kept at -70°C until mindin was analyzed. Serum concentrations of mindin were measured in 56 patients (mean age 68 years, range 45–82 years) with prostate cancer, and in the control group consisting of 29 healthy men (mean age 64 years, range 55–78 years). Informed consent was obtained for all of the individuals included in the study. The characteristics of the patients and healthy subjects are summarized in *Table 1*.

Serum mindin levels were assayed using commercially available immunoassay technique ELISA kit (Cusabio, WuHan, China). The analytic characteristics of the diagnostic kit were as follows: detection limit 0.78 ng/mL and working range 3.12–200 ng/mL, with a mean coefficient of variation (CV)=11%. The patients were previously clinically investigated and classified according to the European Association of Urology (EAU) guidelines (10). For further investigation, the patients were divided into two groups according to the severity of the disease. The first group consisted of patients with less severe tumours (stages 1 and 2), the second group consisted of patients with severe tumours (stages 3 and 4).

Results

Differences between subgroups were tested for statistical significance by the nonparametric Mann-Whitney test. The value of $P < 0.005$ was considered statistically significant. Receiving operation analysis (ROC) was used to investigate the diagnostic efficiency. The analysis of variance was used to evaluate the relationship of mindin levels with the age and PSA levels. Statistical software MedCalc version: 18.02.01 (Ostende, Belgium) was used for statistical analysis.

Serum levels of mindin in patients with prostate cancer were significantly decreased in the control group expressed as medians (2.12 ng/mL vs 0.78 ng/mL, $P = 0.0007$, Mann-Whitney test, *Figure 1*). Diagnostic efficiency of serum mindin expressed as AUC calculated from the ROC analysis was 0.705 (specificity=73 % and sensitivity=64 %) (*Figure 2*). Serum levels of mindin in patients with less severe tumours (stages 1 and 2) and severe tumours (stages 3 and 4) were significantly decreased compared with the control group as well. ($P = 0.0037$, One-way analysis of variance, *Figure 3*).

Discussion

The results of the pilot study confirm the relevant role of mindin in the prostate cancer diagnosis. We proved that the serum levels of mindin were significantly decreased in patients with prostate cancer compared with healthy individuals. These results differ from the results of other studies showing elevated mindin levels in patients with prostate cancer

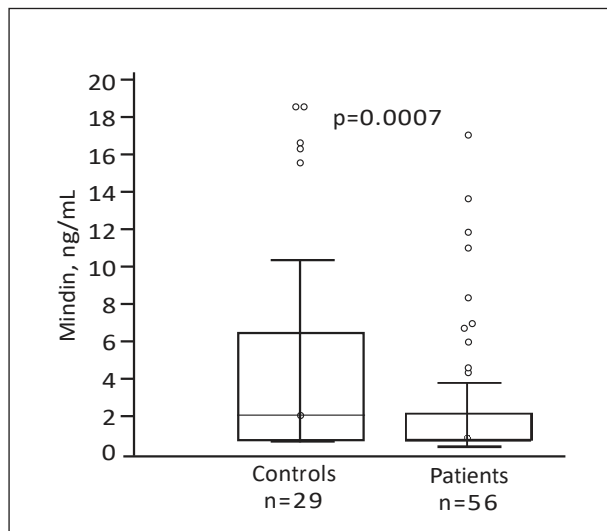


Figure 1 Serum mindin levels in healthy individuals and patients with prostate cancer. Results are expressed as Box-and-whisker plots with medians (IQR 25. – 75. percentile).

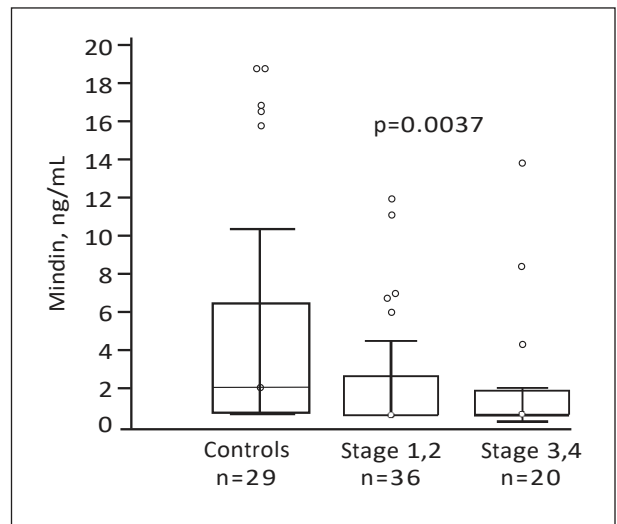


Figure 3 Serum mindin levels in patients with less severe and severe prostate cancer tumours. Results are expressed as Box-and-whisker plots with medians (IQR 25. – 75. percentile)
 Stage 1, 2 – patients with less severe tumours
 Stage 3, 4 – patients with severe tumours

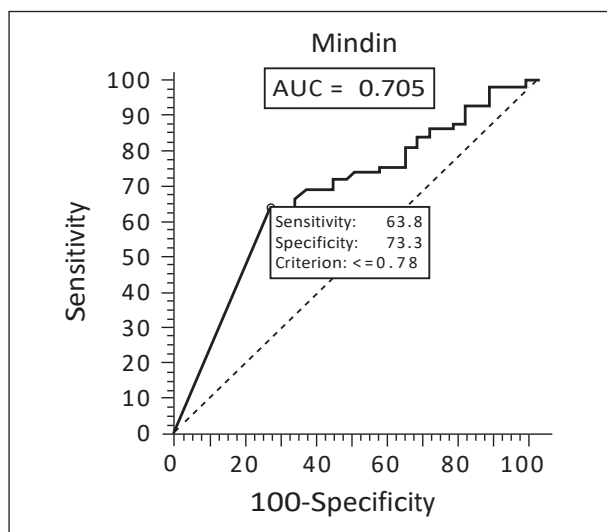


Figure 2 Diagnostic efficiency of mindin.

(11, 12, 14). Presented results are in concordance with the study of Wang et al. (8) showing significantly decreased serum mindin in tumour lesions of patients compared with adjacent control tissues by colon, lung, gastric, oesophageal, and breast cancer. Wang et al. (8) mentioned an important role of EGR-1, which directly regulates mindin expression at the transcriptional level, and this regulates both mindin mRNA and protein expression in vitro to further define EGR-1 mediated regulation of mindin expression. The relationship between EGR-1 expression and prostate cancer was previously mentioned. Gregg et al. showed that EGR-1 is downregulated in patients with prostate carcinoma according to their clinical

considerations (18 of 20 patients in pT2 and pT3 clinical stages showed normal or downregulated EGR-1 expression) (12). Contrary to the previously mentioned reports of Wang et al. (8) and Gregg et al. (12), studies of other authors showed that in patients with prostate cancer EGR-1 stimulates tumour cell growth and its expression level increases with the degree of malignancy (11, 13). This seems to be specific to the prostate tumour cells because, in mammary and lung tumours as well as most normal tissues, EGR-1 expression is low. This contradictory findings regarding EGR-1 expression confirmed the bivalent role of EGR-1 either as a tumour suppressor or oncogene with respect to EGR-1 regulation and the degree of the malignancy.

Our results show good diagnostic sensitivity of mindin with AUC of 0.705. We prove that serum mindin levels are not dependent on the PSA levels, and the age ($P=0.42$, analysis of variance). The serum levels of mindin differ in patients with prostate cancer. We found significant differences between serum mindin concentrations related to the staging of cancer. The patients with less severe tumours belonging to stages 1 and 2 had lower serum mindin level than patients with severe tumours stages 3 and 4 as shown in *Figure 3*. These results correlate with the results found in other cancer types presented by Wang et al., (8) who presented decreased levels in patients with less severe tumours belonging to stages 1 and 2. In our study, the difference between patients in stages 1 and 2 and 3 and 4 is not significant.

The presented study is a pilot study, and thus the results need to be confirmed on a large number of samples. Various enzymatic immunoassays used for

mindin evaluation show discrepant results, as indicated in the presented study and the study of Wang et al. (8) with similar results, which are different from the study of Luccarelli et al. (14). This finding seems to indicate that the concentrations of mindin in patients with prostate cancer are assay dependent.

The concentration of mindin is decreased in patients with prostate cancer. Mindin concentration is not related to the age and PSA levels. Mindin appears to be a promising diagnostic marker useful in the diagnosis of prostate cancer.

References

- Geryk E, Dítě P, Kozel J, [tampach R, Kubířek P, Odehnal J: Other primary neoplasm in patients with prostate cancer in comparison of its incidence, mortality and prevalence. *Onkologie* 2010, 4(2): 89–94.
- Ayyildiz SN, Ayyildiz A: PSA, PSA derivatives, proPSA and prostate health index in the diagnosis of prostate cancer. *Turk J Urol* 2014, 40(2): 82–88.
- Bickers B, Aukim-Hastie C: New molecular biomarkers for the prognosis and management of prostate cancer—the post PSA era. *Anticancer Res* 2009, 29(8): 3289–98.
- Buhmeida A, Pyrhonen S, Laato M, Collan Y: Prognostic factors in prostate cancer. *Diagn Pathol* 2006, 1:4.
- Shariat SF, Canto EI, Kattan MW, Slawin KM: Beyond prostate-specific antigen: new serologic biomarkers for improved diagnosis and management of prostate cancer. *Rev Urol* 2004, 6(2): 58–72.
- Pryor MB, Schellhammer PF: The pursuit of prostate cancer in patients with a rising prostate-specific antigen and multiple negative transrectal ultrasound-guided prostate biopsies. *Clin Prostate Cancer* 2002, 1(3): 172–6.
- Jeli} M, Mandi} A, Kladar N, Sudji J, Bo`in B, Srdjenovi} B. Lipid Peroxidation, Antioxidative Defense and Level of 8-Hydroxy-2-Deoxyguanosine in Cervical Cancer Patients. *J Med Biochem* 2018; 37: 336–45.
- Wang LF, Liu YS, Yang B, Li P, Cheng XS, Xiao CX, Liu JJ, Li S, Ren JL, Guleng B: The extracellular matrix protein mindin attenuates colon cancer progression by blocking angiogenesis via Egr-1-mediated regulation. *Oncogene* 2018, 37(5): 601–15.
- Qian X, Li C, Pang B, Xue M, Wang J, Zhou J: Spondin-2 (SPON2), a more prostate-cancer-specific diagnostic biomarker. *PLoS One* 2012, 7(5): e37225.
- Heidenreich A, Aus G, Bolla M, Joniau S, Matveev VB, Schmid HP, Zattoni F: EAU guidelines on prostate cancer. *Eur Urol* 2008, 53(1): 68–80.
- Gitenay D, Baron VT: Is EGR1 a potential target for prostate cancer therapy? *Future Oncol* 2009, 5(7): 993–1003.
- Gregg JL, Brown KE, Mintz EM, Piontkivska H, Fraizer GC: Analysis of gene expression in prostate cancer epithelial and interstitial stromal cells using laser capture microdissection. *BMC Cancer* 2010 10:165.
- Parra E, Ortega A, Saenz L: Down-regulation of Egr-1 by siRNA inhibits growth of human prostate carcinoma cell line PC-3. *Oncol Rep* 2009, 22(6): 1513–8.
- Lucarelli G, Rutigliano M, Bettocchi C, Palazzo S, Vavallo A, Galleggiante V, Trabucco S, Di Clemente D, Selvaggi FP, Battaglia M et al.: Spondin-2, a secreted extracellular matrix protein, is a novel diagnostic biomarker for prostate cancer. *J Urol* 2013, 190(6): 2271–7.

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Conflict of interest statement

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