

In this work, we present the characteristics of *kha1 $\Delta$*  strains in the background of various multiple mutations in genes encoding alkali-metal-cation transporters. Two main phenotype manifestations of the *kha1* deletion were growth defect on high external pH and hygromycin sensitivity. The correlation between these phenotypes and the *kha1* deletion was confirmed by plasmid complementation. Fluorescence microscopy of GFP-tagged *Kha1p* showed that this antiporter is localized preferentially intracellularly (in contrast to the plasma-membrane  $\text{Na}^+/\text{H}^+$  antiporter *Nha1p*). Based on these findings, *Kha1p* is probably not localized in plasma membrane and does not mediate efflux of alkali metal cations from cells (as published before in Ramírez et al., 1998), but is important for the regulation of intracellular cation homeostasis and optimal pH control, similarly as the *Nhx1p*. The *kha1* deletion phenotypes were complemented by heterologous expression of a plant antiporter *AtChx17*, showing that the proteins *AtChx17* and *SeKha1* could have similar function and that *S. cerevisiae kha1* deletion mutants could serve for heterologous expression and characterization of some plant transporters in yeast, especially those localized intracellularly.

We also showed that the presence of the Tok1 channel strongly influences membrane potential: Deletion of the *TOK1* gene results in significant plasma membrane depolarization, whereas strains overexpressing the *TOK1* gene are hyperpolarized. A decrease in membrane potential is now the second known phenotype (besides the changes in  $\text{Cs}^+$  tolerance) of the *tok1* deletion in *S. cerevisiae* cells. We proved that plasma membrane potential is not the only parameter determining the hygromycin B sensitivity of yeast cells and that the role of intracellular transporters in protecting against its toxic effects must also be considered.